

UDC: 616.12-005.6:616.127-008.313]-092:612.33.015

[https://doi.org/10.32345/USMYJ.1\(144\).2024.18-25](https://doi.org/10.32345/USMYJ.1(144).2024.18-25)

Received: September 21, 2023

Accepted: February 07, 2024

## Fecal short chain fatty acids role in atrial fibrillation paroxysm pathogenesis through coronary artery disease patients

**Melnychuk Iryna, Lyzogub Viktor**

Bogomolets National Medical University, Kyiv, Ukraine

**Address for correspondence:**

Melnychuk Iryna

E-mail: [ira.merkulova45@gmail.com](mailto:ira.merkulova45@gmail.com)

**Abstract:** gut microbiota composition and its metabolites is an essential part of human health. Short chain fatty acids (SCFA) are known gut microbiota metabolites. Lack of them is common for dyslipidemia and inflammatory changes. But their role in atrial fibrillation (AF) and coronary artery disease (CAD) pathogenesis is still uninvestigated. The aim: to estimate the fecal short chain fatty acids changes in patients with atrial fibrillation paroxysm and coronary artery disease and found their connections with known cardiometabolic risk factors. Materials and methods: 300 patients were investigated. We divided them into 3 groups: I group – 149 CAD patients without rhythm disorders, II group – 124 patients with CAD and AF paroxysm and control group (CG) – 27 patients without CAD and arrhythmias. Fecal SCFA was checked by gas chromatography with mass electron detection. Results: Fecal SCFA changes in patients with AF paroxysm and CAD were found in our investigation. Isocaproic and isobutyric fecal acids appears in CAD and AF patients' samples in comparison with control group. In the patients with AF and CAD significant increasing of valeric (1128,43%) and decreasing butyric (78,75%), isovaleric (56,29%), caprylic (99,21%) acids, medium chain fatty acids (95,54%) and unsaturated fatty acids (38,76%) levels was revealed in comparison with CAD patients without arrhythmias ( $P < 0,05$ ). The largest amount of correlations was between total amount of SCFA, medium chain fatty acids (total amount = 7), butyric acid (total number = 6) and cardiometabolic risk factors ( $P < 0,05$ ). The acceptable role of total amount of short chain fatty acids ( $AUC = 0.7907$ ) and butyric acid ( $AUC = 0.7127$ ) in AF paroxysm occurrence in CAD patients was proven by ROC-analysis. Conclusions: SCFA-synthesis violations were revealed in patients with atrial fibrillation paroxysm and coronary artery disease. To propose the new ways of gut microbiota and cardiometabolic risk factors correction will be interesting for future investigations.

**Keywords:** [Coronary Artery Disease](#), [Atrial Fibrillation](#), [Fatty Acids](#), [Cardiometabolic Risk Factors](#), [Patients](#).

### Introduction

Atrial fibrillation (AF) is the most common arrhythmia in the world. By the latest data more than one third European population over 55 suffers from AF. In projection increase of AF will be growth each year. One of the know AF risk factor is coronary artery disease (CAD) (Hindricks G. et all., 2020). CAD is the most

common cardiovascular pathology in the world. Its spreading is also increasing (Knuuti J. et all., 2020). One of explanation of such demographic picture is that CAD and AF have a lot of identical risk factors: dyslipidemia, smoking, obesity, diabetes, obstructive sleep apnea, inflammatory diseases, sedentary way of life, etc. (Hindricks G. et all., 2020; Knuuti J. et all., 2020). Moreover,

CAD and AF presence worsening clinical picture and prognosis of each other, increase risks of cardiovascular events (Michniewicz E. et al., 2018).

The majority of CAD and AF risk factors are pathogenetically linked with gut microbiota condition and its metabolites. By the literature data, inflammatory and metabolic disorders are strongly linked with dysbiosis presence. Gut microbiota violations can act at host organism by its metabolites, which rises in blood flow due to increasing intestinal barrier permeability and their production lesions. Gut microbiota metabolites are trimethylamine (TMA), trimethylamine-N-oxide (TMAO), lipopolysaccharide, bile acids and short chain fatty acids (SCFA) (Malesza IJ et al., 2021; Scheithauer TPM et al., 2020). TMA, TMAO are well known gut microbiota metabolites. Their role in AF paroxysm and CAD pathogenesis is widely discussed nowadays (Gatarek P et al., 2021). Lipopolysaccharide (endotoxin) is also presented as AF paroxysm risk factor (Zhang Y et al., 2022). Elevated levels of circulated bile acids are a well-known AF risk factor (Michelle SW et al., 2019).

SCFA is a crucial gut microbiota metabolite in regulating host immune homeostasis. SCFA are synthesized in human intestine from dietary fibers through fermentation by microorganisms. Deficiency of SCFA content leads for cardiovascular and metabolic disorders by the latest evidence. Some laboratory findings suggested the importance of SCFA role in AF paroxysm development (Lizogub V.G. et al., 2019; Ling Z. et al., 2022), but any clinical investigations. Moreover, it is no evidence about SCFA role in AF paroxysm in CAD patients.

### **Aim**

The aim to estimate the fecal short chain fatty acids changes in patients with atrial fibrillation paroxysm and coronary artery disease and found their connections with known cardiometabolic risk factors.

### **Materials and methods**

300 patients were investigated. We divided them into 3 groups: I group – 149 CAD patients without rhythm disorders, II group – 124 patients with CAD and AF paroxysm and control group (CG) – 27 patients without

CAD and arrhythmias. All diagnosis was established according current European Society of Cardiology guidelines (Hindricks G. et al., 2020; Knuuti J. et al., 2020). Diagnosis CAD was confirmed by history of coronary arteries stenotic changes during invasive coronarography. AF paroxysm was checked by resting 12 leads electrocardiography. Criteria of exclusion were: valvular atrial fibrillation, heart failure (HF) from Class III to IV (by New York Heart Association), reported malignancies, chronic kidney disease (Glomerular Filtration Rate, GFR < 60 mL/min), thyroid pathology, inflammatory bowel disease, irritable bowel syndrome, pregnancy, taking probiotics and antibiotics for a month before the study. There were no vegetarians or vegans among the examined. All patient had HF stage B or C (McDonagh T. et al., 2023). The study was conducted at the base and was approved by the ethical commission of the Kiev City Clinical Hospital No.12. Informed consent was obtained from all subjects in accordance with the Declaration of Helsinki and ethical commission submission. Baseline characteristics of investigated groups are performed in table 1.

Fecal SCFA was checked by gas chromatography with mass electron detection. We determined nine fatty acids in the collected samples – acetic acid (C2:0), propionic acid (C3:0), butyric acid (C4:0), isobutyric acid (C4:1), valeric acid (C5:0), isovaleric acid (C5:1), caproic acid (C6:0), isocaproic acid (C6:1) and caprylic acid (C8:0). These fatty acids include saturated (SFA) – acetic (C2:0), propionic (C3:0), butyric (C4:0), valeric (C5:0), caproic (C6:0), caprylic (C8:0) acids; and unsaturated (USFA) – isobutyric (C4:1), isovaleric (C5:1), isocaproic (C6:1) acids. Middle chain fatty acids (MCFA) include caproic acid (C6:0), isocaproic acid (C6:1) and caprylic acid (C8:0) (Michelle SW et al., 2019). Cardiometabolic risk factors which was explored are: total cholesterol (TC), tryglicerides (TG), low density lipoproteins (LDL), high density lipoproteins (HDL), lipoproteins  $\alpha$  (Lp $\alpha$ ), apolipoprotein A1 (ApoA1), apolipoprotein B (ApoB), C-reactive protein (CRP), interleukine – 6 (IL-6), TMA and TMAO (Lizogub V.G. et al., 2019, Li J.J. et al., 2022). Results were presented as mean  $\pm$  standard error

**Table I.** Baseline characteristics of investigated groups, mean ± standard error

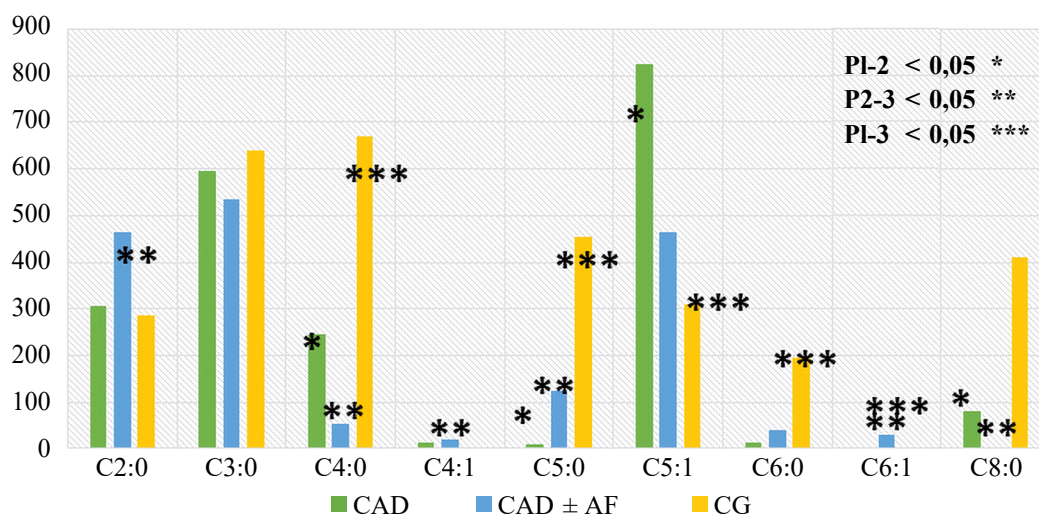
Characteristic /group	I group	II group	CG	P1-2	P2-3	P1-3
Age (years)	67.71±3.90	67.96±0.94	56.25±2.18	P>0.05	P>0.05	P>0.05
Men (%)	48.99	47.97	48.15	P>0.05	P>0.05	P>0.05
History of myocardial infarction (%)	30,87	26,02	0	P>0,05	P<0,05	P<0,05
History of stroke (%)	8,72	8,13	0	P>0,05	P<0,05	P<0,05
Diabetes mellitus	18.12	14.63	0	P>0,05	P<0,05	P<0,05
Obesity	8.84	12.0	0	P>0,05	P<0,05	P<0,05
BMI (kg/m <sup>2</sup> )	27.02±0.33	26.93±0.43	28.12±2.10	P>0.05	P>0.05	P>0.05
Smoking (%)	51.01	41.46	40.74	P>0.05	P>0.05	P>0.05
Uric acid (mmol/l)	380.5±28.16	404.9±36.11	310.2±29.12	P>0.05	P<0.05	P<0.05
Total bilirubin (mmol/l)	11.3±0.09	12.4±0.08	11.7±0.11	P>0.05	P>0.05	P>0.05
GFR (ml/min)	62.03±2.31	67.73±1.98	84.01±5.48	P>0.05	P<0.05	P<0.05

or [95% confidence interval (CI)] for continuous variables or as a number for categorical variables. Variables distribution for normality were checked by the Pearson criterion. Data were compared by Scheffe's or Dann multiple comparison methods depends with two critical regions for variables distribution respectively; Spearman's rank correlation coefficient was detected. ROC-curves with area under ROC-curve (AUC) calculation for gut microbiota component and their combinations were built for I and II groups (Faizi et al., 2023; Mandrekar JN, 2010). All calculations were done in MATLAB R2014a (License number 271828).

**Results**

Fecal SCFA composition was studied in all investigated groups. Total amount of fecal SCFA

in I (71,13%) and II (41,89%) patients' groups is significantly decreased in comparison with CG. Also, increasing of acetic (62,35%) and decreasing butyric (92,21%), valeric (72,36%), caprylic (99,84%) acids levels in 2 group in comparison with CG was found. In the I group was found significant arising of isovaleric (62,35%) and abundance of butyric (63,36%), valeric (97,75%), caproic (93,39%) acids in comparison with CG. In the II group significant increasing of valeric (1128,43%) and decreasing butyric (78,75%), isovaleric (56,29%), caprylic (99,21%) acids levels was revealed in comparison with I group. Isocaproic and isobutyric fecal acids were absent into the CG samples, but they appeared in I and II groups patients' tests. Results are shown in figure 1.



**Fig. 1.** Fecal short chain fatty acids in investigated groups, mg/g



Moreover, in II group was found significant increase of USFA (485,44%) and decrease of MCFA (66,04%) levels in comparison with CG. In I group was found significant growth of USFA (258,54%) and decreasing of MCFA (98,49%) in comparison with CG. Also, in II group was significant decreasing MCFA (95,54%) and USFA (38,76%) levels in comparison with I group. Results are shown in figure 2.

Secondary, lipid profile, inflammatory markers and TMA, TMAO levels of investigated groups were evaluated. In the I and II group was a significant increasing of TC (32.64% and 43.06% respectively), TG (80.36% and

55.36% respectively), LDL (70.78% and 72.73% respectively), Lpα (41.17% and 54.95% respectively), ApoB (85.12% and 140.50% respectively), CRP (136.26% and 232.97% respectively), IL-6 (65.22% and 103.11% respectively), TMA (22.50% and 42.25% respectively), TMAO (50.00% and 136.31% respectively) and decreasing HDL (16.09% and 29.31% respectively) compared with CG. In the II group significant increase of ApoB (29.91%), CRP (40.93%), IL-6 (22.93%), TMA (16.13%), TMAO (57.54%) levels were detected in comparison with a I group. Results are shown in table 2.

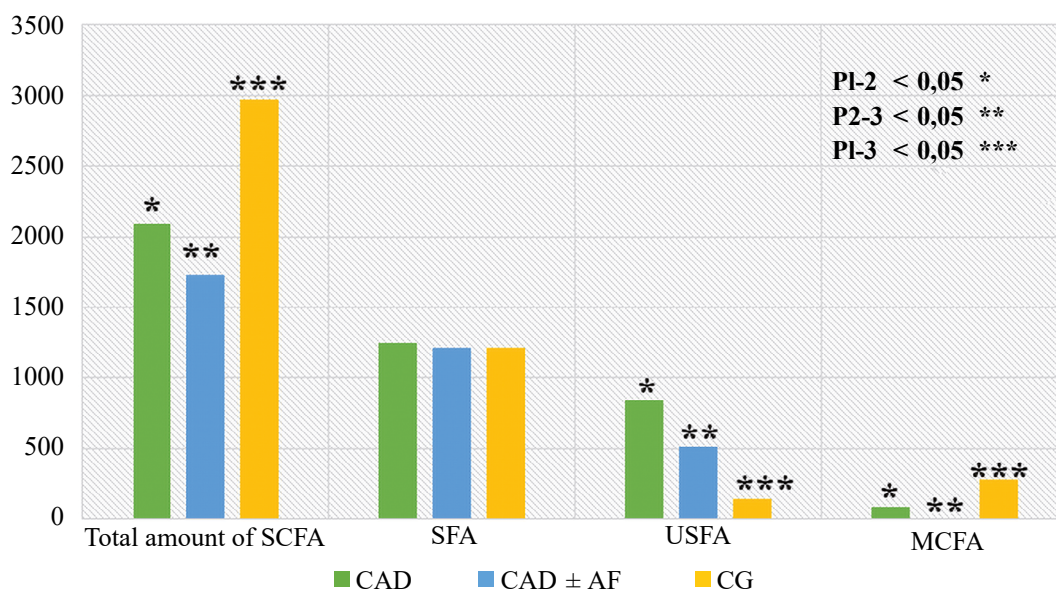
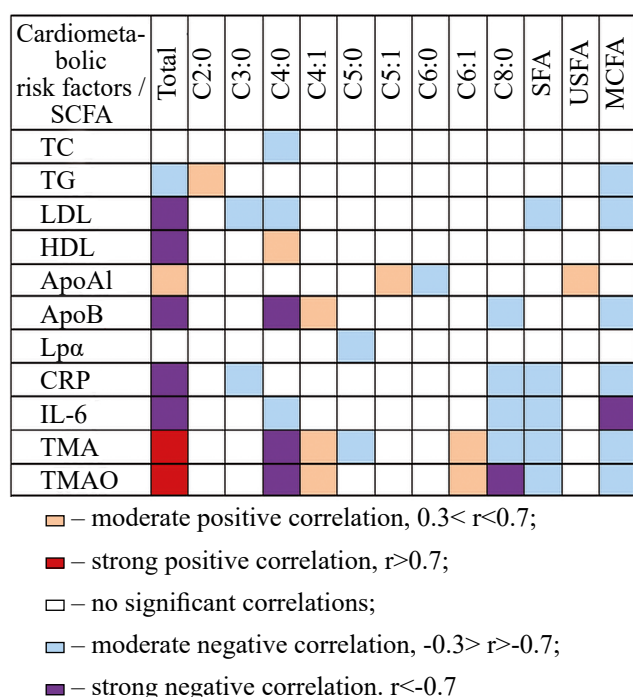


Fig. 2. Fecal short chain fatty acids in investigated groups, mg/g

Table II. Cardiometabolic risk factors of investigated groups, mean ± standard error

Characteristic /group	I group	II group	CG	P1-2	P2-3	P1-3
TC (mmol/l)	5.73±0.37	6.18±0.31	4.32±0.21	P>0.05	P<0.05	P<0.05
TG (mmol/l)	2.02±0.18	1.74±0.14	1.12±0,09	P>0.05	P<0.05	P<0.05
LDL (mmol/l)	2.63±0.29	2.66±0.24	1.54±0.11	P>0.05	P<0.05	P<0.05
HDL (mmol/l)	1.46±0.13	1.23±0.14	1.74±0.12	P>0.05	P<0.05	P<0.05
Lpα (mg/dl)	22.53±1.26	24.73±1.48	15.96±1.23	P>0.05	P<0.05	P<0.05
Apo A1 (g/l)	2.02±0.16	2.34±0.27	1.62±0.09	P>0.05	P>0.05	P>0.05
Apo B (g/l)	2.24±0.19	2.91±0.13	1.21±0.18	P<0.05	P<0.05	P<0.05
CRP, mg/l	2.15±0.20	3.03±0.19	0.91±0.12	P<0.05	P<0.05	P<0.05
IL-6, pg/ml	2.66±0.16	3.27±0.16	1.61±0.09	P<0.05	P<0.05	P<0.05
TMA, mmol/l	21,89±0,45	25,42±0,37	17,87±0,50	P<0.05	P<0.05	P<0.05
TMAO, mmol/l	2,52±0,11	3,97±0,13	1,68±0,11	P<0.05	P<0.05	P<0.05

Further, heatmap correlation matrices was generated between lipid profile and fecal SCFA levels. The largest amount of correlations was checked between fecal SCFA composition and such clinical characteristics as TMAO (total number = 8), TMA (total number = 7) and CRP (total number = 6) levels. At the same time, the highest amount of correlations was between total amount of SCFA (total number = 9), MCFA (total amount = 7), butyric acid (total number = 6) and cardiometabolic risk factors. It is shown in figure 3.



**Fig. 3.** Fecal SCFA levels correlations with clinical and laboratory changes,  $P < 0.05$

ROC-analysis was done for each SCFA for better understanding their diagnostic value in pathogenesis of AF paroxysm in CAD patients. We calculated AUC for each sign. Results are shown in table 3.

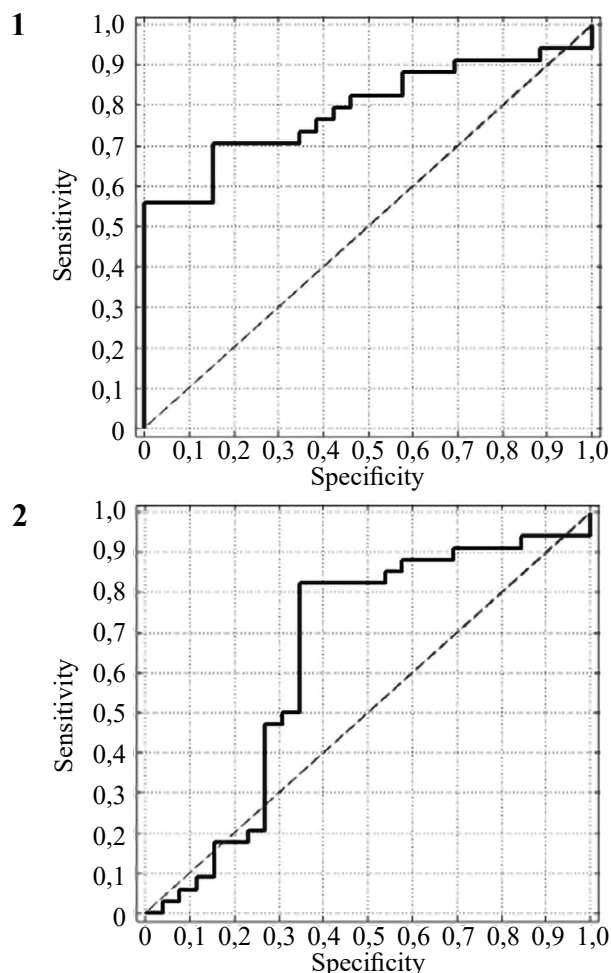
AUC was more then 0,7 (acceptable) was found in three sings: total amount of SCFA (AUC = 0.7907) and butyric acid (AUC = 0.7727). They are shown in figure 4.

**Discussion**

SHFA are mainly produced by such probiotics as Roseburia, Eubacterium rectale, Blautia and Ruminococcus from dietary plant polysaccharides. Lack of this species leads to

**Table 3.** Area under ROC-curve to each bacterium in patients I and II group,  $P < 0.05$

Gut microbiota	AUC
Total amount of SCFA	<b>0.7907</b>
C2:0	0.5577
C3:0	0.6437
C4:0	<b>0.7127</b>
C4:1	0.6572
C5:0	0.6855
C5:1	0.6917
C6:0	0.6538
C6:1	0.6527
C8:0	0.6782
SFA	0.6461
USFA	0.5213
MCFA	0.6988



**Fig. 4.** ROC-curve analysis for SCFA in I and II groups,  $P < 0.05$ : 1 – total amount of SCFA, AUC = 0.7907; 2 – butyric acid, AUC = 0.7127.

impaired intestinal mucosal barrier function and increased bacterial endotoxin secretion, which were directly correlated with host metabolic and inflammatory disorders (Xiao S. et al., 2019; Patterson E. et al., 2016). SCFA cardioprotective effect is based on modulation T regulatory cell amount. Also, decreasing of total SCFA concentration is common foe diabetes, arterial hypertension, nonalcoholic steatohepatitis formation (Mandrekar JN, 2010). So, in general SCFA concentration is very important for gut microbiota stability (Lizogub V.G. et al., 2019).

Different SCFA are different in their role and tissue distribution. Butyrate is the main energy resource for colonocyte. Propionate activate liver gluconeogenesis. In the host organism they available to inhibit host histone deacetylases, which takes part in the protein's synthesis (Coppola S. et al., 2021). Furthermore, different microbes produce different SCFA. For example, butyrate is mainly produced by Gram positive microorganisms, as Firmicutes, acetate and propionate – by Gram negative microorganisms, as Bacteroides. The type of SCFA production depends of different factors, including diet, gut microbiota composition, species evolution and colonic environment. After colonocytes absorb SCFA and they coming into blood flow. These SCFA can be used for carbohydrates and lipids synthesis of like cytokines for metabolism regulation. SCFA are able to activating brown adipose tissue, regulating liver mitochondrial function, maintaining body energy homeostasis, controlling appetite and sleep (He, J. et al., 2020).

SCFA plays essential role in lipids metabolism. By the latest data butyrate increase oxidation of fatty acids in brown adipose tissue, reduce the size of adipose cells, regulate activity of transcription factors, what leads for decreasing levels of triglycerides and fatty acids (He, J. et al., 2020; Schoeler M. et al., 2019). All of this confirmed the importance of SCFA and especially butyric acid in CAD pathogenesis. By literature data total amount SCFA decrease is associated with CAD presence (Lizogub V.G. et al., 2019).

Role of SCFA in AF pathogenesis is still undoubted, but there is a multiplicity of data about their role in AF risk factors pathogenesis (Hu, T. et al., 2022; Gawałko M. et al., 2022).

So, SCFA further investigations are promising for new AF risk factors investigation and correction.

### Conclusions

Fecal short chain fatty acids changes in patients with atrial fibrillation paroxysm and coronary artery disease were found in our investigation:

1. Isocaproic and isobutyric fecal acids appears in coronary artery disease and atrial fibrillation patients' samples in comparison with control group.
2. In the patients with atrial fibrillation and coronary artery disease significant increasing of valeric (1128,43%) and decreasing butyric (78,75%), isovaleric (56,29%), caprylic (99,21%) acids, medium chain fatty acids (95,54%) and unsaturated fatty acids (38,76%) levels was revealed in comparison with coronary artery disease patients without arrhythmias ( $P < 0,05$ ).
3. The largest amount of correlations was between total amount of short chain fatty acids, medium chain fatty acids (total amount = 7), butyric acid (total number = 6) and cardiometabolic risk factors ( $P < 0,05$ ).
4. The acceptable role of total amount of short chain fatty acids (AUC = 0.7907) and butyric acid (AUC = 0.7727) in AF paroxysm occurrence in CAD patients was proven by ROC-analysis.

### Perspectives of subsequent scientific research

To propose the new ways of gut microbiota and cardiometabolic risk factors correction will be interesting for future investigations.

### Financing

This study did not receive external funding. The study was done according the department scientific research work "Changes in protein, carbohydrate and lipid metabolism in patients with coronary heart disease and arterial hypertension with heart rhythm disorders, possibilities of drug correction" 2021-2023 (state registration number 0121U108875)

### Conflicts of Interest

it is no conflict of interest to declare.

### Consent to publication

Informed consent was obtained from all subjects in accordance with the Declaration of Helsinki and ethical commission submission.



**ORCID ID and Autor contributions**[0000-0002-0659-1476](https://orcid.org/0000-0002-0659-1476)

(A,B,C,D,E)

Melnychuk Iryna

[0000-0003-3603-7342](https://orcid.org/0000-0003-3603-7342)

(A,E,F) Lyzogub

Viktor

A – Research concept and design, B – Collection and/or assembly of data, C – Data analysis and interpretation, D – Writing the article, E – Critical revision of the article, F – Final approval of article

**REFERENCES**

Coppola S, Avagliano C, Calignano A, Berni Canani R. The Protective Role of Butyrate against Obesity and Obesity-Related Diseases. *Molecules*. 2021 Jan 28;26(3):682. doi: 10.3390/molecules26030682. PMID: 33525625; PMCID: PMC7865491.

Faizi, Nafis, and Yasir Alvi. *Biostatistics Manual for Health Research*. Elsevier, 15 Jan. 2023.

Gatarek P, Kaluzna-Czaplinska J. Trimethylamine N-oxide (TMAO) in human health. *EXCLI J*. 2021 Feb 11;20:301-319. doi: 10.17179/excli2020-3239. PMID: 33746664; PMCID: PMC7975634.

He, J., Zhang, P., Shen, L., Niu, L., Tan, Y., Chen, L., Zhao, Y., Bai, L., Hao, X., Li, X., Zhang, S., & Zhu, L. (2020). Short-Chain Fatty Acids and Their Association with Signalling Pathways in Inflammation, Glucose and Lipid Metabolism. *International journal of molecular sciences*, 21(17), 6356. <https://doi.org/10.3390/ijms21176356>

Hindricks G., Potpara T., Dagres N. et al. ESC Guidelines for the diagnosis and management of atrial fibrillation developed in collaboration with the European Association for Cardio-Thoracic Surgery (EACTS). *European Heart Journal*. 2020. Vol. 42. P. 373498. doi:10.1093/eurheartj/ehaa612

Hu, T., Wu, Q., Yao, Q., Jiang, K., Yu, J., & Tang, Q. (2022). Short-chain fatty acid metabolism and multiple effects on cardiovascular diseases. *Ageing research reviews*, 81, 101706. <https://doi.org/10.1016/j.arr.2022.101706>

Knuuti J., Wijns W., Saraste A. et al. ESC Guidelines for the diagnosis and management of chronic coronary syndromes. *European Heart Journal*. 2020. Vol. 41. P. 407477. doi:10.1093/eurheartj/ehz425

Li J.J., Liu H.H., Li S. Landscape of cardiometabolic risk factors in Chinese population: a narrative review. *Cardiovasc Diabetol*. 2022. Vol. 21(1). P.113. doi: 10.1186/s12933-022-01551-3.

Lizogub V.G., Kramarova V.N., Melnychuk I.O. The role of gut microbiota changes in the pathogenesis of heart disease. *Zaporizkiy medical journal*. 2019. V. 21, No 5 (116). P. 672–678. doi: 10.14739 / 2310-1210.2019.5.179462

Malesza IJ, Malesza M, Walkowiak J, Mussin N, Walkowiak D, Aringazina R, Bartkowiak-Wieczorek J, Mądry E. High-Fat, Western-Style Diet, Systemic Inflammation, and Gut Microbiota: A Narrative Review. *Cells*. 2021 Nov 14;10(11):3164. doi: 10.3390/cells10113164. PMID: 34831387; PMCID: PMC8619527.

Mandrekar JN Receiver Operating Characteristic Curve in Diagnostic Test Assessment. *Journal of Thoracic Oncology*. 2010;5(9);1315-1316.

Michelle SW., Xiang, Laurence Macia et al. Fatty Acids, Gut Bacteria, and Immune Cell Function The Molecular Nutrition of Fats, 2019

Michniewicz E., Młodawska E., Lopatowska P., Tomaszuk-Kazberuk A., & Malyszko J. Patients with atrial fibrillation and coronary artery disease - Double trouble. *Advances in medical sciences*. 2018. Vol. 63(1). P. 30–35. <https://doi.org/10.1016/j.advms.2017.06.005>

Scheithauer TPM, Rampanelli E, Nieuwdorp M, Vallance BA, Verchere CB, van Raalte DH, Herrema H. Gut Microbiota as a Trigger for Metabolic Inflammation in Obesity and Type 2 Diabetes. *Front Immunol*. 2020 Oct 16;11:571731. doi: 10.3389/fimmu.2020.571731. PMID: 33178196; PMCID: PMC7596417.

Xiao S, Zhang Z, Chen M, Zou J, Jiang S, Qian D, Duan J. Xiexin Tang ameliorates dyslipidemia in high-fat diet-induced obese rats via elevating gut microbiota-derived short chain fatty acids production and adjusting energy metabolism. *J Ethnopharmacol*. 2019 Sep 15;241:112032. doi: 10.1016/j.jep.2019.112032. Epub 2019 Jun 18. PMID: 31220598.

Zhang Y, Zhang S, Li B, Luo Y, Gong Y, Jin X, Zhang J, Zhou Y, Zhuo X, Wang Z, Zhao X, Han X, Gao Y, Yu H, Liang D, Zhao S, Sun D, Wang D, Xu W, Qu G, Bo W, Li D, Wu Y, Li Y. Gut microbiota dysbiosis promotes age-related atrial fibrillation by lipopolysaccharide and glucose-induced activation of NLRP3-inflammasome. *Cardiovasc Res*. 2022 Feb 21;118(3):785-797. doi: 10.1093/cvr/cvab114. PMID: 33757127.

Theresa A McDonagh, Marco Metra, Marianna Adamo, Roy S Gardner, Andreas Baumbach, Michael Böhm, Haran Burri, Javed Butler, Jelena Čelutkienė, Ovidiu Chioncel, John G F Cleland, Maria Generosa Crespo-Leiro, Dimitrios Farmakis, Martine Gilard, Stephane Heymans, Arno W Hoes, Tiny Jaarsma, Ewa A Jankowska, Mitja Lainscak, Carolyn S P Lam, Alexander R Lyon, John J V McMurray, Alexandre Mebazaa, Richard Mindham, Claudio Muneretto, Massimo Francesco Piepoli, Susanna Price, Giuseppe M C Rosano, Frank Ruschitzka, Anne Kathrine Skibelund, ESC Scientific Document Group, 2023 Focused Update of the 2021 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: Developed by the task force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC) With the special contribution of the Heart Failure Association (HFA) of the ESC, *European Heart Journal*, Volume 44, Issue 37, 1 October 2023, Pages 3627–3639, <https://doi.org/10.1093/eurheartj/ehad195>

Patterson E, Ryan PM, Cryan JF, Dinan TG, Ross RP, Fitzgerald GF, Stanton C. Gut microbiota, obesity and diabetes. *Postgrad Med J.* 2016 May;92(1087):286-300. doi: 10.1136/postgradmedj-2015-133285. Epub 2016 Feb 24. PMID: 26912499.

Schoeler M, Caesar R. Dietary lipids, gut microbiota and lipid metabolism. *Rev Endocr Metab Disord.* 2019 Dec;20(4):461-472. doi: 10.1007/s11154-019-09512-0. PMID: 31707624; PMCID: PMC6938793.

Gawałko M, Agbaedeng TA, Saljic A, Müller DN, Wilck N, Schnabel R, Penders J, Rienstra M, van Gelder I, Jespersen T, Schotten U, Crijns HJGM, Kalman JM, Sanders P, Nattel S, Dobrev D, Linz D. Gut microbiota, dysbiosis and atrial fibrillation. Arrhythmogenic mechanisms and potential clinical implications. *Cardiovasc Res.* 2022 Aug 24;118(11):2415-2427. doi: 10.1093/cvr/cvab292. PMID: 34550344; PMCID: PMC9400433.

Ling Z, Liu X, Cheng Y, Yan X, Wu S. Gut microbiota and aging. *Crit Rev Food Sci Nutr.* 2022;62(13):3509-3534. doi: 10.1080/10408398.2020.1867054. Epub 2020 Dec 30. PMID: 33377391.

## Роль фекальних коротколанцюгових жирних кислот в патогенезі пароксизму фібриляції передсердь у хворих на ішемічну хворобу серця

Мельничук Ірина, Лизогуб Віктор

Національний медичний університет імені О.О. Богомольця, Київ, Україна

### Address for correspondence:

Melnychuk Iryna

E-mail: [ira.merkulova45@gmail.com](mailto:ira.merkulova45@gmail.com)

**Анотація:** склад кишкової мікробіоти та її метаболіти є важливою складовою здоров'я людини. Коротколанцюгові жирні кислоти (КЛЖК) є відомими метаболітами кишкової мікробіоти. Їх нестача характерна для дисліпідемії та запальних змін. Але їх роль у патогенезі фібриляції передсердь (ФП) та ішемічної хвороби серця (ІХС) досі не вивчена. Мета: оцінити зміни коротколанцюгових жирних кислот у фекаліях у пацієнтів з пароксизмом фібриляції передсердь та ішемічною хворобою серця та встановити їх зв'язок з відомими кардіометаболічними факторами ризику. Матеріали і методи: Обстежено 300 хворих. Ми розподілили їх на 3 групи: I група – 149 хворих на ІХС без порушень ритму, II група – 124 пацієнти з ІХС та пароксизмом ФП та контрольна група (КГ) – 27 пацієнтів без ІХС та аритмій. Фекальні КЛЖК визначали за допомогою газової хроматографії з мас електронною детекцією. Результати: У нашому дослідженні виявлено зміни вмісту КЛЖК у калі у пацієнтів з пароксизмом ФП та ІХС. Ізокапронова та ізомасляна фекальні кислоти виявляються у зразках хворих на ІХС та ФП порівняно з КГ. У хворих на ФП та ІХС суттєво підвищувався вміст валеріанової (1128,43%) та знижувався масляної (78,75%), ізовалеріанової (56,29%), каприлової (99,21%) кислот, середньоланцюгових жирних кислот (95,54%) та ненасичених жирних кислот (38,76%) порівняно з хворими на ІХС без аритмій ( $P < 0,05$ ). Найбільша кількість кореляцій була між загальною кількістю КЛЖК, середньоланцюгових жирних кислот (загальна кількість = 7), масляної кислоти (загальна кількість = 6) і кардіометаболічними факторами ризику ( $P < 0,05$ ). ROC-аналізом доведено важливу роль загальної кількості КЛЖК ( $AUC = 0.7907$ ) та масляної кислоти ( $AUC = 0.7127$ ) у виникненні пароксизму ФП у хворих на ІХС. Висновки: У хворих на пароксизм фібриляції передсердь та ішемічну хворобу серця виявлено порушення синтезу коротколанцюгових жирних кислот. Запропонувати нові способи корекції кишкової мікробіоти та кардіометаболічних факторів ризику буде цікаво для майбутніх досліджень.

**Ключові слова:** ішемічна хвороба серця, фібриляція передсердь, жирні кислоти, кардіометаболічні фактори ризику.



Copyright: © 2024 by the authors; licensee USMYJ, Kyiv, Ukraine.

This article is an open access article distributed under the terms

and conditions of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>).